

Δήμητρα Ντάφου

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A Simplified History of Life on Earth



How is the human genome organised?

- 5% coding and rest of it *junk* (repetitive DNA).
- Nuclear and mitochondrial
- You are 99.99% similar to your neighbor

Κατανομή γονιδίων στον άνθρωπο



Gene Regulatory Mechanisms

- Transcriptional Mechanisms
 - Type of promoters & RNA polymerase
 - Control of Transcription
 - Constitutive
 - Inducible
 - Repressible
 - Transcription Factors and TFBS
- Translational Mechanisms
 - Micro RNAs (miRNAs and RITS complexes)
 - Translational control
 - mRNA degradation
 - Promoter activation
 - Silencer RNAs (siRNAs & RISC complexes) degrading mRNA
- Epigenetic Mechanisms
 - Chromatin remodeling
 - Histone modifications (acetylation, phosphorylation, methylation ...)
 - DNA methylation

Gene Expression Regulatory Network



Non-coding RNA: the formerly known as "junk"

NC-RNAs compose majority of transcription in complex genomes



Nc-RNAS- a bit of a 'Dark Matter' Function



RNA interference first discovered in Petunias Called PTGS, for

"Post Transcriptional Gene Silencing"



Color changes can be induced by RNAi, or PTGS..

> Post transciptional gene silencing

Small (21-23 nts) RNA duplexes, with the same sequence as in the silenced gene, were identified as being responsible for knocking down expression



So what other organisms can do this thing called PTGS?

"Post Transcriptional Gene Silencing"













C. Elegans grow on agar dishes with *E. coli* bacteria as a source of food.

If they eat *E. coli* expressing dsRNA molecules...this creates specific knock-down of gene expression!



In *C. elegans* the siRNA effects can be amplified making the silencing quite stable

This does not appear to happen in mammalian cells

(RISC = RNAi Induced Silencing Complex; RdRP = RNA dependent RNA polymerase)

RNA sets the standard

Thomas Tuschl

One way of finding out what genes do is to inactivate them, and to study the effects, in 'model' organisms. That has now been done for many thousands of worm genes in two large-scale analyses.



Figure 1 Gene screening by double-stranded-RNA-mediated interference (RNAi). Kamath *et al.*¹ and Ashrafi *et al.*² used the following technique to silence the expression of 16,757 genes individually in *Caenorhabditis elegans*. **a**, DNA molecules (plasmids) encoding a double-stranded RNA (dsRNA) of choice are inserted into *Escherichia coli* bacteria. Incubation with isopropylthio- β -galactoside (IPTG) induces production of the dsRNA. **b**, Worms at the latest larval stage are placed on a lawn of *E. coli*, and allowed to feed. Several adult worms are then placed onto new plates seeded with the same bacteria to lay eggs. The offspring are monitored for embryonic death and post-embryonic phenotypes, such as slow larval growth or movement disorders.

http://www.nature.com/cgi-taf/DynaPage.taf?file=/nature/journal/v421/n6920/full/421220a_fs.html&content_filetype=pdf



19,757 genes

16,757 have been inactivated by RNAi

10% display spontaneous phenotype; this 10% is enriched for conserved genes



19,757 genes

16,757 knock down mutants were screened for body fat content

305 knock downs had increased body fat, 112 genes had decreased...

new targets for obesity?

NOBEL PRIZE IN PHYSIOLOGY 2006



The Nobel Prize in Physiology or Medicine 2006

"for their discovery of RNA interference - gene silencing by doublestranded RNA"





Andrew Z. Fire

1/2 of the prize

USA

Stanford University School of Medicine Stanford, CA, USA

Photo: R. Carlin/UMMAS

1/2 of the prize USA University of Massachusetts Medical School Worcester, MA, USA

Craig C. Mello

Cho WC. MicroRNAs in cancer - from research to therapy. Biochim Biophys Acta - Rev Cancer 2010;1805(2):209-217.

Potent and specific genetic interference by double-stranded RNA in Caenorhabditis elegans

Andrew Fire*, SiQun Xu*, Mary K. Montgomery*, Steven A. Kostas*†, Samuel E. Driver‡ & Craig C. Mello‡

C. elegans

NATURE VOL 391 19 FEBRUARY 1998



So...what about RNAi in mammalian cells...

May 2001...the first report...

letters to nature

Nature 411, 494 - 498 (2001); doi:10.1038/35078107

Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells

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RNA interference (RNAi) is the process of sequence-specific, post-transcriptional gene silencing in animals and plants, initiated by double-stranded RNA (dsRNA) that is homologous in sequence to the silenced gene. The mediators of sequence-specific messenger RNA degradation are 21- and 22-nucleotide small interfering RNAs (siRNAs) generated by ribonuclease III cleavage from longer dsRNAs. Here we show that 21-nucleotide siRNA duplexes specifically suppress expression of endogenous and heterologous genes in different mammalian cell lines, including human embryonic kidney (293) and HeLa cells. Therefore, 21-nucleotide siRNA duplexes provide a new tool for studying gene function in mammalian cells and may eventually be used as gene-specific therapeutics.

How does RNAi work in mammalian cells?

RNAi works post-transcriptionally......

in key two steps!



Dicer contains two RNAse III domains



siRNAs have a defined structure

19 nt duplex



step two:

the antisense strand of the siRNA guides cleavage



Tuschl, 2002

RNAi silencing complex

may be associated with translating ribosomes

active RNAse enzyme not yet identified

 may participate in endogenous pathways that silence genes via translational repression

HOW MICRORNAS ARE PRODUCED?



• Active complex: miRISC

Nature Reviews MCB, 2008

Biogenesis of miRNA



1. The initial gene transcript is called primary miRNA (pri-miRNA)

2.In the cell nucleus, these hairpin-loop molecules are cut to form double-stranded precursor miRNA (pre-miRNA)

3. The pre-miRNA is transported to the cytoplasm. There, it is further cut to form a functional mature miRNA (mature miRNA molecules are about 22 nucleotides long)

4. The mature miRNA first binds with a molecule called the RNA interference silencing complex, or RISC

5. Then the miRNA binds with its target messenger RNA (mRNA), thereby blocking its translation or prompting its degradation



siRNA mediated degradation of mRNA

versus

miRNA mediated inhibition of mRNA translation

How microRNAs work?

- Partial complementarity with 3'UTR regions (position 2 to 8 critical)
- Cooperative effect
- Abrogate protein synthesis



One microRNA may regulate up to 100 different genes!

miRNA Binding



miRNAs Inhibit Translation by Inducing Ribosome Drop-Off



ribosome drop-off

Petersen et al. (2006) Molecular Cell 21: 533-542.

miRNA Expression Results in Temporal and Spatial Reciprocity with Target Expression

a Temporal reciprocity



b Spatial reciprocity



Genomic Organization of miRNA Genes



•Exonic miRNAs - non-coding (or in alternatively spliced exons) Zhao Y, Srivastava D, TIBS 32:189,2007

Precise expression profile





Zhao et al, Nature 2005

Pena et al, Nature Methods 2009

Loss of microRNA leads to fatality

Loss of miR-1-2 leads to overproduction of muscle cells

Ventura et al, Cell 2008

Zhao et al, Cell 2007





Loss of miR-17-92 cluster is embryonic lethal

Mechanisms linking microRNAs with disease

• Genomic alterations of microRNAs

- Chromosome deletion, amplification, and translocation
- Single nucleotide polymorphism of miRNA or miRNA targets
- Alterations on the expression of levels of miRNAs
 - Transcriptional control: transcription factor, enhancer, repressor
 - Epigenetic modification: DNA methylation, histone acetylation
- Alteration on the processes of microRNA biogenesis

microRNA: a new class of biomarkers



small noncoding RNAs that regulate gene expression by binding complementary sequences of target mRNAs and inducing their degradation or translational repression Evolutionary conserved

One miRNA has multiple targets



microRNAs linked to diseases

• Viral infections

- Viruses encodes microRNAs that target viral mRNAs to regulate various stages of the viral life cycle
- Viral microRNAs suppress expression of specific host genes
- Viral infections induce expression of host microRNAs that inhibits expression of cellular genes
- Upon viral infections, host cells express specific microRNA that suppress viral mRNA expression
- Cardiac, immune, neurological and metabolic disorders

MicroRNAs and Cancer

Different expression pro between healthy and ca tissue samples



Lu et al, Nature 2005

MicroRNAs in cancer therapy



Decrease in tumour mass

Kota et al. Cell, 2009





Predictive Modelling

Permit risk stratification.

Customize treatment

Less extensive surgery

Rational drug selection

Monitoring response to therapy

Predictive or Diagnostic Modelling

Use of one or more biomarkers to determine prognosis or response to treatment beyond usual clinical criteria

- Tissue based
- Serum or urinary based
- Cellular based

Why are microRNAS such good biomarkers?

- •Extremely stable in fluids as well as on formalin-fixed paraffin-embedded tissue
- •Expression profile correlates well between fresh and formalin-fixed paraffin-embedded samples
- Resistant to degradation

Unique microRNA profile in lung cancer diagnosis and prognosis

 A univariate Cox proportional hazard regression model with a global permutation test indicated that expression of the miRNAs *hsa-mir-155* and *hsa-let-7a-2* were related to adenocarcinoma patient outcome

1.0 hsa-mir-155 low expression 0.8 Survival rate 0.6 0.4 hsa-mir-155 high expression 0.2 p=0.006 0.0 40 20 60 80 100 120 Months 1.0 0.8 hsa-let7a-2 high expression





Yanaihara N, et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. Cancer Cell 2006, 9:189-198.

Study Design for Biomarker Validation



Clinical Obsevation (Clinomics, Phenomics, etc.) Clinical Variables and Phenotypes (e.g. obesity, hypertension) Associated with Cardiotoxicity Data from Clinical Studies, Electronic Health Records, and Other Big Data

	Genomics	Epigenomics	Transcriptomics	Proteomics	Metabolomics	Immunolomics
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Targets	DNA (e.g. SNP or WGS)	Chromatin accessibility Chromatin structure DMA methylation	mRNA Non-coding RNA (e.g. miRNA, piRNA, IncRNA)	Secreted and intracellualr proteins	Endogenous circulating metabolites Xenobiotics	immune cells Cytokines
Detection Technology	NGS (DNA-seq) Microarrays	ATAC-seq ChiP-seq Methyl-seq	NGS (RNA-seq) Microamays	Affinity-based (e.g. antibody aptamers) Mass spectrometry	Mass spectrometry Nuclear magnetic resonance spectroscopy	Immuno-seq CyTOF Single-cell omics Proteomics
Applications	1. Genetic variants to predict susceptibility 2. Polygenic risk score 3. CHIP	1. Epigenetic footprint to predict susceptibility 2. Epigenetic modification caused by cardiotoxicity	1. Transcriptomic signatures and/or gene target:/pathways caused by cardiotoxicity 2. Circulating non-coding RNAs predictive of cardiotoxicity	1. Protein biomarker 2. Protein-based risk score to predict cardiotoxicity	1. Metabolites correlated with cardiotoxicity- related metabolic impairment	1. Distinct immune cell populations or cell phenotypes associated with cardiotoxicity 2. Discovery of cytokine patterns

Integrated Omics, Computational Biology, Machine Learning

Novel Biomarker Discovery



microRNA Biomarker discovery workflow and panel selection options







MicroRNA profiling Platforms

Conventional cancer treatment:

Personalized cancer treatment:



