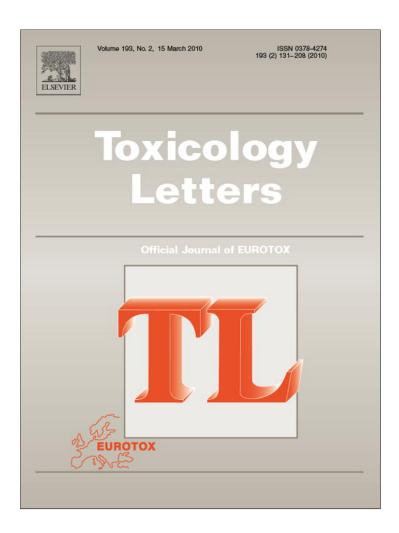
Provided for non-commercial research and education use. Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

http://www.elsevier.com/copyright

# Author's personal copy

Toxicology Letters 193 (2010) 131-137



Contents lists available at ScienceDirect

# **Toxicology Letters**

journal homepage: www.elsevier.com/locate/toxlet



## Mini review

# Environmental factors and genetic susceptibility promote urinary bladder cancer

Dimitrios Volanis a,c,1, Tanya Kadiyska a,1, Alex Galanis b,1, Dimitrios Delakas a, Stella Logothetic, Vassilis Zoumpourlis c,\*

- <sup>a</sup> Department of Urology, "Asklipieio" General Hospital, Voula, Athens, Greece
- <sup>b</sup> Department of Molecular Biology and Genetics, Democritus University of Thrace, Dragana, 68100 Alexandroupolis, Greece
- <sup>c</sup> Unit of Biomedical Applications, Institute of Biological Research and Biotechnology, National Hellenic Research Foundation, 48 Vas. Constantinou Ave, 116 35 Athens, Greece

### ARTICLE INFO

#### Article history: Received 30 October 2009 Received in revised form 20 December 2009 Accepted 21 December 2009 Available online 4 January 2010

Keywords: Bladder cancer Environmental risk factor Genetic polymorphism miRNAs Molecular profiling

### ABSTRACT

Cancer of the urinary bladder is the second most common malignancy of the genitourinary tract, currently accounting for up to 5% of all newly diagnosed tumours in the western world. Urinary bladder carcinogenesis seems to develop from the interaction of environmental exposure and genetic susceptibility. Smoking, specific industrial chemicals, dietary nitrates and arsenic represent the most important exogenous risk factors. Chromosomal abnormalities, silencing of certain genes by abnormal methylation of their promoter region, alterations in tumour suppressor genes and proto-oncogenes that induce uncontrolled cell proliferation and reduced apoptosis, are molecular mechanisms that have been reported in bladder carcinogenesis. In this article, we discuss the environmental risk factors of bladder cancer and we review the genetic and epigenetic alterations, including aberrant DNA methylation and deregulation of microRNAs expression. We also discuss the role of p53 and retinoblastoma suppressor genes in disease progression. Finally, we present recent reports on the use of molecular profiling to predict disease stage and grade and direct targeted therapy.

© 2010 Elsevier Ireland Ltd. All rights reserved.

## Contents

1.	Introduction	131
	Environmental risk factors	
3.	Genetic and epigenetic alterations	133
	3.1. Chromosomal aberrations	133
	3.2. Aberrant DNA methylation	
	3.3. Altered microRNAs expression	134
	3.4. Loss of p53 and retinoblastoma gene function	134
	Molecular profiling	
5.	Conclusion	136
	Conflict of interest	136
	Acknowledgments	136
	References	136

## 1. Introduction

Cancer of the urinary bladder is the 5th most common cancer in the Western world, responsible for approximately 14,000 cancerrelated deaths in the United States each year (Jemal et al., 2009). Bladder cancer (BC) can occur at any age; however it is generally a disease of the middle-aged or elderly. The median ages at diagnosis are 69 years in males and 71 in females and it is about 3 times more common in men than in women. The increased life expectancy of the Western population has led to a concomitant rise in the incidence of bladder cancer during the last 20 years, currently accounting for up to 5% of all new cancers. Ninety-eight percent of all bladder cancers are epithelial malignancies, with the vast majority being transitional cell carcinomas (TCC). Other rare histologic types include squamous-cell carcinomas, adenocarcinomas, and nonepithelial cancers. The most common finding in

<sup>\*</sup> Corresponding author at: Laboratory of Biomedical Applications Unit, National Hellenic Research Foundation, Institute of Biological Research and Biotechnology, 48, Vas Constantinou Ave, 116 35 Athens, Pagrati, Greece. Tel.: +30 210 727 3730; fax: +30 210 727 3677.

E-mail address: vzub@eie.gr (V. Zoumpourlis).

Equal contribution.

bladder cancer is microscopic or gross haematuria. However, the clinical manifestations are associated with stage and thus range from asymptomatic disease to uremic coma. For instance weight loss, skeletal pain and oedema of the lower extremities can rarely be found in non-muscle invasive tumours, whereas they represent common findings in locally advanced and metastatic disease. At presentation 75–85% of patients have tumours confined to the urothelial mucosa (stage Ta) or lamina propria (stage T1). Nevertheless, more than 60% of these tumours will recur at least once and progress to less differentiated or invasive neoplasms ( $\geq$ T2) in 10–15% of all cases. The remaining patients present with muscle invasive tumours (stages T2–T4) or with *de novo* metastatic disease (5%), having less favourable prognosis.

Bladder cancer follows the general concept of multi-step carcinogenesis and it is likely that multiple lesions on the DNA of target cells are required for malignant transformation. Moreover, similar carcinogens may facilitate the development of different genetic alterations due to the microenvironment of transitional epithelium. Bladder carcinogenesis is thought to develop from the interaction of environmental exposures and genetic susceptibility. Herein, we present the environmental risk factors of bladder cancer and we review the genetic and epigenetic alterations and the key signalling molecules involved. Furthermore, we present the molecular profiling, a novel technology based on simultaneous determination of multiple molecular markers by using microarrays and focus on its application to predict disease stage, progression and clinical outcome.

### 2. Environmental risk factors

The most important environmental factors associated with bladder cancer are listed on Table 1. Tobacco smoke is considered the most important exogenous risk factor for bladder cancer (Stewart et al., 2008). It is estimated that 50% of TCCs are directly attributable to tobacco smoke. Arylamines and more than 60 other carcinogens present in tobacco smoke are associated with DNA adduct formation and alterations in tumour suppressor genes expression (Wallerand et al., 2005). Smokers present with higher grade and stage tumours, ultimately having higher bladder cancerspecific mortality compared to non-smokers. Cessation of smoking leads to a 40% reduction of bladder cancer risk within one year, however the risk remains increased for as many as 25 years.

Occupational exposure accounts for approximately 20% of all bladder cancer cases. Aromatic amines (e.g. benzidine,  $\beta$ napthylamine, and 4-aminobiphenyl) present in aniline dyes and many other industrial chemicals have been reported to be causally related to bladder cancer (Rouissi et al., 2009; Golka et al., 2004). Occupational urinary bladder cancer has been observed in painters, varnishers, hairdressers, textile, rubber and leather industry workers. The latency period between exposure and clinical presentation is related to cumulative dose and ranges from 30 to 50 years. Perchloroethylene is a chlorinated hydrocarbon found mostly in dry-cleaning solvents that has been associated with the development of bladder cancer. A review of the epidemiologic literature demonstrated a substantial higher risk for the development of bladder cancer in laundry and dry-cleaning workers (Mundt et al., 2003). Similarly, polycyclic aromatic hydrocarbons have been related with the development of respiratory and urinary tract cancers (Bosetti et al., 2007).

The uncontrolled use of nitrogen-based fertilizers and pesticides has led to nitrate contamination of soil and groundwater. The relation between dietary nitrates and the development of TCC is controversial. A recent study conducted in Taiwan residents demonstrated a significant positive relationship (Chiu et al., 2007), whereas a cohort study in Dutch population did not support an association between dietary nitrates and bladder cancer (Zeegers et al., 2006). Many other epidemiologic studies relate the high levels of chloride and arsenic in drinking water with increased risk of developing bladder cancer (Villanueva et al., 2003; Chiou et al., 2001). The presumable mechanisms for arsenic carcinogenesis are DNA methylation changes, aberrant gene expression, altered cell metabolism and oxidative stress (Huang et al., 2004).

The association between pelvic radiotherapy and the development of bladder cancer has been well documented in the past (Kaldor et al., 1995). Moreover, studies performed in the Ukraine population after the Chernobyl accident, linked the chronic exposure to persistent low-dose of ionizing radiation (IR) to the activation of specific molecular pathways and the subsequent initiation of urinary bladder carcinogenesis (Romanenko et al., 2000; 2002; 2003; 2009). More specifically, it has been demonstrated that long-term, low-dose of IR induces oxidative stress, causing DNA damage (Romanenko et al., 2000) significant activation of DNA repair enzymes (base and nucleotide excision repair), 8-hydroxy-2'deoxyguanosine (8-OHdG), 8-oxoguanine-

**Table 1** Environmental risk factors for the development of bladder cancer.

Environmental factor	Comment	Reference
Smoking	50% of cases	Stewart et al. (2008)
	Higher cancer-specific mortality	Wallerand et al. (2005)
Aromatic amines	Aniline dyes and other industrial chemicals	Rouissi et al. (2009)
		Golka et al. (2004)
Ionizing radiation	Pelvic radiotherapy	Romanenko et al. (2009)
	Radio-contaminated areas	Kaldor et al. (1995)
Arsenic	Contamination of water	Huang et al. (2004)
		Chiou et al. (2001)
Dietary nitrites/nitrates	Fertilizers and pesticides	Chiu et al. (2007)
		Zeegers et al. (2006)
Chloride	Chlorinated water	Villanueva et al. (2003)
	Weak correlation	
Chlorinated hydrocarbons	Dry-cleaning solvents	Mundt et al. (2003)
	Contamination of soil	
Polycyclic aromatic hydrocarbons	Aluminium production, coal gasification, coke production, tar and tar-ralated products	Bosetti et al. (2007)
Alkylating agents	Cyclophosphamide	Knight et al. (2004)
	Carcinogenic metabolite (acrolein)	Tanguay et al. (2003)
Coal	Coal miners	Lopez-Abente et al. (2006)
Schistosomiasis	Schistosoma haematobium	Gouda et al. (2007)
	Endemic in Africa, Asia and South America	Fedewa et al. (2009)
	Squamous-cell carcinoma	

DNA-glycosylase (OGG1), apurinic/apyrimidinic endonuclease 1 (APE1) and xenoderma pigmentosum A (XPA) (Romanenko et al., 2002), overexpression of p38 mitogen-activated protein (MAP) kinase and cytoplasmic retention of NF-kB (Romanenko et al., 2003), as well as epigenetic alterations, such as aberrant DNA methylation (see Section 3.2) (Franco et al., 2008).

A remarkable observation is that individuals with seemingly equal exposure to environmental carcinogens vary enormously in their risk of developing cancer. This is attributed probably to the fact that the carcinogenic detoxification system and the DNA repair capacity vary within the human population. Much interest is concentrated on the significance of genetic polymorphism in the glutathione S-transferase (GST) supergene family involved in cellular metabolism and in detoxification of cytotoxic and carcinogenic products. In a recent study, involving 731 bladder cancer patients and 740 control subjects, GSTM1, GSTT1 and GSTP1 null genotypes were associated with a 19-48% increase in odds ratio (OR) of bladder cancer (Yuan et al., 2008). Similarly, slow and rapid acetylators of aromatic amines demonstrate variable susceptibility to TCC. Current evidence suggests a relationship between N-acetyltransferase 2 (NAT-2) and more pronounced urothelial damage (Rouissi et al., 2009; Song et al., 2009). In addition, a study designed to evaluate the importance of genetic instability in developing bladder cancer, strongly suggested that individuals better able to resist DNA damage are less susceptible to TCC (Schabath et al., 2003). Accordingly, polymorphisms of the X-ray repair cross-complementing 1 (XRCC1) and the human oxoguanine glycosylase 1 (hOGG1) genes that encode enzymes of the base excision repair (BER) mechanism for repairing DNA damage, are risk factors of bladder cancer (Arizono et al., 2008). In addition, single nucleotide polymorphisms (SNPs) in the 8-oxoG DNA-glycosylase (OGG1), the poly(ADPribose) polymerase family member 1 (PARP1) and the major gap filling polymerase-beta (POLB) BER genes also showed significant associations with bladder cancer risk (Figueroa et al., 2007). Some cytokines, such as the vascular endothelial growth-factor (VEGF) and the tumour necrosis factor-alpha (TNF-alpha) which are crucial for angiogenesis, local invasion and metastasis, are encoded by genes that are also polymorphic. Recently, there has been an association between the aggressiveness of TCC and certain VEGF and TNF-alpha genotypes (Kim et al., 2005). Accordingly, a large-scale evaluation of candidate cancer genes has identified three SNPs in the promoter region of VEGF that were associated with increased risk for bladder cancer (García-Closas et al., 2007).

# 3. Genetic and epigenetic alterations

## 3.1. Chromosomal aberrations

The characteristic behaviour of bladder cancer has given the opportunity to investigate the genomic alterations present in newly diagnosed and recurrent tumours, of various grades and stages. Numerous structural and numerical changes have been identified, some representing an early step, while others seem to be related with progression. Two general observations that came from studies based on comparative genomic hybridization (CGH) and single nucleotide polymorphism (SNP) arrays are that tumours of different stages demonstrate different chromosomal aberrations and that higher rate of chromosomal aberrations are present in muscle invasive tumours (stages  $\geq$ T2) compared to superficial tumours (stages Ta, T1) (Blaveri et al., 2005; Koed et al., 2005). Moreover, it seems that gains and amplifications generally predominate over deletions in advanced-stage tumours. The above mentioned aberrations could lead to fusion products and possibly to altered expression of oncogenes or tumour suppressor genes (see Section 3.4).

The most frequent chromosomal aberrations found in all grades and stages of papillary and solid TCC, is loss of heterozygosity (LOH) at chromosome 9 (Hirao et al., 2005). Deletions of chromosome 9 have also been demonstrated in histological normal, hyperplastic and dysplastic urothelium. These findings support the hypothesis that aberrations of chromosome 9 probably represent an early, or even the initial event in urinary bladder carcinogenesis (Abraham et al., 2007). In most of the cases the chromosome's long arm is affected (9q22, 9q32-33, 9q34). According to Knudson's hypothesis, the above observations suggest that one or more tumour suppressor genes could be located on the long arm of chromosome 9. Candidate genes include the ptch1, the tsc1 (already implicated in other cancers) and the dbc1. The ptch1 gene, also known as the Gorlin Syndrome gene, is found within a small region of deletion at 9q22 and is a tumour suppressor of the hedgehog pathway (Aboulkassim et al., 2003). The tuberous sclerosis complex gene 1 (tsc1) which is located at 9q34 encodes hamartin, a protein that together with tuberin negatively regulate the mTOR (mammalian target of rapamycin) pathway. Germline mutations of tsc1 have been associated with tuberous sclerosis and familial hamartoma syndrome. Finally, the *dbc1* gene, at 9q33, probably exerts non-apoptotic cell death and stall in the G1 phase (Lopez-Beltran et al., 2008).

Deletions at the short arm of chromosome 9 have been also found in TCC of the urinary bladder. Studies are focused on cdkn2a at 9p21, a tumour suppressor most frequently inactivated by homozygous deletion (Chapman et al., 2005). The cdkn2a locus encodes two proteins, p16 and p14<sup>ARF</sup>, both of which are considered key cell cycle regulators. Other chromosomal aberrations found in early-stage tumours include deletions of 8p, 11p and gains of 1q and 17q (Qin et al., 2006).

As mentioned above, the progression of a superficial bladder tumour to muscle invasive disease is associated with the occurrence of a wide spectrum of genetic alterations. This results in major genetic divergence even in synchronous, adjacent tumours. Yet, strong genetic similarities have been documented among invasive TCCs irrespectively of the depth of invasion. The above observation suggests that invasion of the lamina propria could be the hallmark of a more aggressive behaviour, with subsequent further local invasion and ultimately formation of distant metastases. In general, the chromosomal aberrations found predominantly in invasive tumours are deletions of 6q, 11p, 18q and gains of 1q, 8q and 17q. Some of these aberrations reflect alterations in known genes (deletions of 17p13 and 13q14 result in inactivation of *p53* and *Rb1* respectively), whereas the significance of others remains vague (Chan et al., 2009).

## 3.2. Aberrant DNA methylation

Many genes have CpG-rich regions that are grouped in clusters known as CpG islands, found within or near the promoter regions. Hypermethylation of these regions results in loss of gene expression and gene silencing. Several tumour suppressor genes associated with TCC of the urinary bladder demonstrate aberrant methylation and subsequent down-regulation and silencing of gene expression. Among them,  $p16^{INK4A}$  is the one most thoroughly studied. This tumour suppressor alts progression from G1 to S phase by inactivating cyclin-dependent kinase 4 and cyclin-dependent kinase 6. Methylation of  $p16^{INK4A}$  has been found in frequencies ranging from 7% to 60% of TCCs of the urinary bladder. On the contrary, methylation of the  $p15^{INK4b}$  gene which is located in the same locus has been reported to occur only in a small subclass of TCCs, suggesting that there are other mechanisms responsible for its inactivation (Brait et al., 2008; Catto et al., 2005).

The methylation status of 10 different genes was examined from 98 bladder cancer samples (Maruyama et al., 2001). According to

the results of this study, four genes were most frequently methylated: cdh1, cdh13, rassf1a and apc. Moreover, methylation of cdh1, a member of the cadherin family encoding a calcium dependent cell to cell adhesion glycoprotein, was associated with shortened survival. It is presumed that loss of cdh function leads to cancer progression by increasing proliferation rate or by promoting local invasion and metastasis. In a second study, both tissue and urine samples were used to examine the methylation status of lama3, lamb3 and lamc2 (Sathyanarayana et al., 2004). These genes encode a family of extracellular matrix glycoproteins (laminins) implicated in cell adhesion, differentiation, migration and signalling. According to the results of this study, there was an association between methylation of these genes and overall poor prognosis. Moreover, the methylation status of specific p53 target genes was determined by performing quantitative methylation-specific real-time PCR (Christoph et al., 2006). Hypermethylation of the promoter region in tumour samples from 110 tumour patients was detected for apoptotic protein-activating factor (APAF-1) (100%), death-associated protein kinase (DAPK-1) (74%) and insulin-like growth-factor-binding protein-3 (IGFBP-3) (66%), but not for Caspase 8 (CASP-8) (3.6%). Importantly, the APAF-1 and IGFBP-3 methylation levels were able to distinguish tumours with higher recurrence risk from low-risk tumours, thus acting as valuable prognostic markers for reappearance and progression of the disease. Similar results were presented in a recent study, performed in 34 patients with TCC of the bladder (Christoph et al., 2007).

To conclude, methylation takes place early in the process of carcinogenesis and can be detected by non-invasive methods in both serum and urine samples. Furthermore, methylation always occurs in the same DNA location and thus it is easier to be detected compared to other genetic biomarkers (e.g. gene mutations). These characteristics render it as a potential diagnostic tool for the early diagnosis of TCC. Compared to conventional urine cytology, the use of methylation markers seems more sensitive and specific especially for low grade tumours, where cancer cells are more cohesive and not readily shed into the urine. The methylation profile of the promoters of several tumour specific genes has been analysed, as described above, providing invaluable information regarding the progress and the outcome of the disease. Nevertheless, further investigation is needed for routine clinical application of methylation markers (Esteller, 2005).

## 3.3. Altered microRNAs expression

MicroRNAs (miRNAs) are about 22 nucleotide long non-protein coding RNAs that regulate gene expression by interacting with messenger RNAs (mRNAs) and induce target mRNA cleavage and translational repression (Tang et al., 2008). Aberrant miRNAs expression, up-regulation or down-regulation, has been linked to the process of tumourigenesis (Giannakakis et al., 2007). Recent studies reported that specific miRNAs play a critical role in TCC development and progression. These findings are presented in Table 2. Firstly, the differential expression of these miRNAs can act as a primary invaluable marker to distinguish cancer from normal tissue samples. Moreover, a miRNA expression profile analysis may provide information regarding the stage of the disease and be indicative for cancer progression and prognosis. miRNAs can act either as tumour suppressors (miR-17-5p, miR-21, miR-126, miR-221), or as oncogenes (miR-26a, miR-29c, miR-30c, miR-30e-5p). Although, their exact target genes, the molecular pathways they are involved and the clinopathological outcome of their abnormal expression, have not yet been fully elucidated, the miRNAs still represent attractive candidates for gene therapy (Adam et al., 2009). One delicate approach that has recently been developed is the design and application of synthetic antisense oligonucleotides that specifically target oncogenic miRNAs and suppress their expres-

**Table 2**Altered expression of miRNAs in bladder cancer.

miRNA	Putative target gene	Reference		
(a) Up-regulation				
miR-10a	FGFR3	Veerla et al. (2009)		
miR-17-5p	c-Myc	Gottardo et al. (2007)		
miR-21	PTEN	Dyrskjøt et al. (2009)		
miR-23a		Gottardo et al. (2007)		
miR-23b		Gottardo et al. (2007)		
miR-26b		Gottardo et al. (2007)		
miR-30-3p	KRT7	Ichimi et al. (2009)		
miR-103-1	HOX	Gottardo et al. (2007)		
miR-125b		Veerla et al. (2009)		
miR-126	EGFL7	Saito et al. (2009)		
miR-129	GALNT1 and SOX4	Dyrskjøt et al. (2009)		
miR-133a	KRT7	Ichimi et al. (2009)		
miR-143	Ras	Lin et al. (2009)		
miR-182		Hanke et al. (in press)		
miR-185		Gottardo et al. (2007)		
miR-199a*	KRT7	Ichimi et al. (2009)		
miR-203		Gottardo et al. (2007)		
miR-205		Gottardo et al. (2007)		
miR-221	p27 <sup>kip</sup>	Gottardo et al. (2007)		
miR-222	p27 <sup>kip</sup>	Veerla et al. (2009)		
miR-223		Gottardo et al. (2007)		
miR-452		Veerla et al. (2009)		
(b) Down-regulation				
miR-7	FGFR3	Veerla et al. (2009)		
miR-26a	707.10	Wang et al. (in press)		
miR-29c		Wang et al. (in press)		
miR-30c		Wang et al. (in press)		
miR-30e-5p		Wang et al. (in press)		
miR-31		Veerla et al. (2009)		
miR-145		Dyrskjøt et al. (2009)		
		, , , , , , , , , , , , , , , , , , , ,		

sion. The use of these antisense oligonucleotides, also termed antagomirs, appears to be effective against miR-221/222 overexpression, impairing the growth of prostate carcinoma xenografts in mice (Mercatelli et al., 2008). Similarly, injection of the antagomir-17-5p could abolish tumour growth in a neuroblastoma mice model (Fontana et al., 2008). These promising findings indicate that the antagomir treatment will to be a powerful therapeutic strategy against cancer in the near future.

# 3.4. Loss of p53 and retinoblastoma gene function

Abnormalities of TP53 have been well described in various malignancies, and germline mutations are associated with the Li-Fraumeni syndrome (Brosh and Rotter, 2009). Alterations of both the P53 and retinoblastoma (RB) molecular pathways play a major role in human bladder carcinogenesis (Shariat et al., 2004). The P53 tumour suppressor protein is encoded by the p53 gene, located at chromosome 17p13.1. The p53 gene has multiple functions, with a central role in tumour suppression by initiating apoptosis or inducing cell arrest at the G1/S phase in response to DNA damage through the induction of p21waf1/cipi. While most bladder cancers exhibit a loss of a single 17p allele, an additional mutation in the second, wild-type allele can inactivate p53 leading to increased nuclear accumulation of the corresponding protein and loss of its tumour suppressor function. The most frequent alteration involves exons 5–11 of p53, especially codons 280 and 285 and the loss of Rb gene function. A recent study performed in 75 non-invasive urinary bladder cancer patients showed that tumour recurrence frequency was 69.4% in patients with wild-type p53, and 88.5% in patients with mutant p53. Moreover, the progression-free survival was significantly shorter in patients with p53 mutations, and the frequency of tumour progression was significantly higher in mutated as compared to wild-type tumours (Ecke et al., 2008). Another mechanism for TP53 gene inactivation is the overexpression of the murine double minutes (mdm2) oncogene. The MDM2 protein promotes the

transcriptional inactivation as well as the proteasomal degradation of P53. MDM2 overexpression has been associated with increasing stage and grade of TCC (Simon et al., 2002).

Alterations in the Rb pathway are also commonly found in the development of invasive urinary bladder cancer. The Rb gene is located at chromosome 13q14 and its product is a nuclear phosphoprotein, which plays a critical role in several pathways involved in urothelial carcinogenesis, including development, differentiation, cell cycle restriction, senescence and apoptosis. The active form (dephosphorylated) of the RB protein binds to the transcription factor E2F, thereby sequestering it. When RB is competitively phosphorylated, it releases E2F, which is able to bind its target genes and promote cell cycle progression. It has been demonstrated the significant association between Rb loss, tumour stage, and tumour grade (Gallucci et al., 2005). In parallel, it has been shown that amplification and overexpression of E2f3 is also associated with increased tumour stage, grade and proliferation index in human bladder cancer (Olsson et al., 2006). Interestingly, a recent study linked Rb inactivation and overexpression of E2f3 isoforms, E2f3a and E2f3b, to amplification of 6p22 in bladder tumours (Hurst et al., 2008). This genomic region contains 4 genes (prl, sox4, E2f3 and cdkal1) and appears to play a critical role in tumour progression.

## 4. Molecular profiling

Several investigations have clearly demonstrated that bladder cancer cannot be managed solely on the basis of pathologic staging. In addition, the ability of the current staging system to estimate the risk of adverse clinical outcome for an individual patient is limited. Similarly, the existing therapeutic methods offer long-term recurrence-free survival, only in a subset of patients (Table 3). Namely, transurethral resection (TUR) can achieve complete resection of non-muscle invasive tumours, but has no influence on the natural history of the disease. Radical cystectomy for T2-T4a tumours offers a 5-year recurrence-free survival in just 68% of patients (Stein et al., 2001), whereas the currently used chemotherapeutic regimens offer long-term disease-free survival in only 15% of patients with metastatic disease. Currently, several new therapies are been tested complementary to the established methods

of treatment in order to increase effectiveness. A recent study demonstrated increased sensitivity to Bacillus Calmette–Guerin (BCG) induced TRAIL release after intravesical H<sub>2</sub>O<sub>2</sub> instillations (White–Gilbertson et al., 2009).

Undoubtedly, a thorough insight into the involved pathways and analysis of the combined effects of several validated markers is required to develop clinically valuable tests and new targeted therapies. As carcinogenesis is a multi-step process, synergistic therapeutic regimens that are aimed at multiple targets are more promising than targeting a single step of a pathway. Over the past years, several members of these pathways have proved to be useful for diagnosis, early detection, therapeutic monitoring and staging of bladder cancer. A big challenge in that field is now the use of cDNA microarrays that allows monitoring simultaneously the expression level of thousands of selected known genes as well as cDNAs representing uncharacterized genes, in one hybridization experiment (Sánchez-Carbayo, 2003).

In an independent study, cDNA microarrays containing 17,842 known genes and expressed sequence tags were used to identify differences in gene expression between normal mucosa, nonmuscle invasive (early-stage), and muscle invasive (late stage) bladder tumours (Sánchez-Carbayo et al., 2003). Differential expression of several genes, including cytokeratin 20, neuropilin-2, p21, and p33ING1 was detected and validated by immunohistochemistry using tissue microarrays. Importantly, the expression patterns of the above genes were significantly associated with pathological stage, tumour grade, and altered Rb expression. Thus, they represent ideal molecular biomarkers of BC progression and potential targets of novel therapeutics. In a similar study, a highdensity oligonucleotide microarrays analysis (59,619 genes and expressed sequence tags) was performed to identify gene clusters associated with BC stage and grade (Dyrskot et al., 2005). Using a cross-validation approach, a 45-gene molecular signature was detected, predicting disease progression and recurrence of pTa tumours. Differentially expressed genes were involved in regulating apoptosis (birc4 and birc6), cell differentiation, and cell cycle progression (mcm7, cdc20). Significantly, this molecular signature has recently been statistically validated in a multicenter analysis of tumour samples from 404 patients diagnosed with BC (Dyrskot et

**Table 3** Treatment of TCC of the urinary bladder.

Stage	Treatment	Comment
CIS	Intravesical BCG+TUR Cystectomy	Standard treatment Patients with incomplete response/recurrence
Ta-T1	TUR TUR + intravesical chemotherapy TUR + intravesical BCG Radical cystectomy	Low-risk patients (followed by a single intravesical dose of chemotherapeutic) Intermediate-risk patients High-risk patients In BCG failure, multiple recurrent, high grade tumours, concomitant CIS
T2-T4a	Radical cystectomy  External beam radiotherapy  Multimodality treatment  Neo-adjuvant chemotherapy + radical cystectomy	Standard treatment 68% recurrence-free survival at 5 years Patients unfit for cystectomy TUR+chemotherapy+radiotherapy (comparable long-term survival) 5-7% improved overall survival
T4b	Radical cystectomy	Palliative
N+	Radical cystectomy + adjuvant chemotherapy	Under debate
M+	Chemotherapy	MVAC (1st line) Carboplatin (unfit patients) Paclitaxel/gemcitabine (2nd line)
Therapeutic pr	rospects	
	Intravesical gene therapy	Non-muscle invasive tumours Limitations in gene delivery to the urothelium
	Blocking VEGF/VEGFR	Bevacizumab (in phase II clinical trial)
	Inhibitors of COX-2	Chemoprevention
	Intravesical H <sub>2</sub> O <sub>2</sub>	Reverses resistance to TRAIL Adjuvant to BCG

al., 2007). Although more similar studies are needed, that include large number of clinical samples and detailed history data, it is evident that molecular profiling of bladder cancer will become an invaluable tool to accurately predict the progression of the disease and the treatment response, as well as to better guide patient treatment and therapy.

### 5. Conclusion

The behaviour of transitional cell carcinoma of the urinary bladder is highly diverse and defined by two separate, but related processes: tumour recurrence and progression. In order to optimize management, patients are categorized into low, intermediate and high-risk groups. The most useful prognostic parameters are tumour grade, stage, architecture (papillary vs. solid), and the synchronous presence of urothelial dysplasia or cancer in situ. Nevertheless, neither recurrence, nor progression rate can be predicted accurately by physiological means alone. It is apparent, that the elucidation of the molecular mechanisms of the disease is vital. In this vein, a significant amount of studies have been performed over the past years, generating important information regarding the genetic and molecular pathways involved. These findings lead to the development of novel prognostic and therapeutic strategies, such as the antagomirs, synthetic antisense oligonucleotides, that specifically target oncogenic miRNAs and suppress their expression and the molecular profiling, a technology based on microarrays for the simultaneous determination of multiple molecular markers or molecular signatures. Further work is required in order to optimise these methods and validate their efficiency. However, as the molecular basis of the disease becomes rapidly understood, there are high expectations for the development of even more powerful diagnostic and therapeutic tools in the near future.

## **Conflict of interest**

The authors declare that there are no conflicts of interest.

## Acknowledgments

This work is supported by the TOK Marie Curie program SUPRAGENE, Cont. No MTKD-CT-2005-029508. T Kadiyska is a fellow in this program.

## References

- Aboulkassim, T., LaRue, H., Lemieux, P., Rousseau, F., Fradet, Y., 2003. Alteration of the PATCHED locus in superficial bladder cancer. Oncogene 22, 2967–2971.
- Abraham, R., Pagano, F., Gomella, L., Baffa, R., 2007. Chromosomal deletions in bladder cancer: shutting down pathways. Front. Biosci. 12, 826–838.
- Adam, L., Zhong, M., Čhoi, W., Qi, W., Nicoloso, M., Arora, A., Calin, G., Wang, H., Siefker-Radtke, A., et al., 2009. miR-200 expression regulates epithelial-to-mesenchymal transition in bladder cancer cells and reverses resistance to epidermal growth factor receptor therapy. Clin. Cancer Res. 15, 5060–5072.Arizono, K., Osada, Y., Kuroda, Y., 2008. DNA repair gene hOGG1 codon 326 and
- Arizono, K., Osada, Y., Kuroda, Y., 2008. DNA repair gene hOGG1 codon 326 and XRCC1 codon 399 polymorphisms and bladder cancer risk in a Japanese population. Jpn. J. Clin. Oncol. 38, 186–191.
- Blaveri, E., Brewer, J., Roydasgupta, R., Fridlyand, J., DeVries, S., Koppie, T., et al.,
   2005. Bladder cancer stage and outcome by array-based comparative genomic hybridization. Clin. Cancer Res. 11, 7012–7022.
   Bosetti, C., Boffetta, P., La Vecchia, C., 2007. Occupational exposures to polycyclic
- Bosetti, C., Boffetta, P., La Vecchia, C., 2007. Occupational exposures to polycyclic aromatic hydrocarbons, and respiratory and urinary tract cancers: a quantitative review to 2005. Ann. Oncol. 18, 431–446.
- Brait, M., Begum, S., Carvalho, A., Dasgupta, S., Vettore, A., Czerniak, B., Caballero, O., et al., 2008. Aberrant promoter methylation of multiple genes during pathogenesis of bladder cancer. Cancer Epidemiol. Biomarkers. Prev. 17, 2786–2794.
- Brosh, R., Rotter, V., 2009. When mutants gain new powers: news from the mutant p53 field. Nat. Rev. Cancer 9, 701–713.
- Catto, J., Azzouzi, A., Rechman, I., Feeley, K., Cross, S., Amira, N., Fromont, G., Sibony, M., et al., 2005. Promoter hypermethylation is associated with tumor location, stage and subsequent progression in transitional cell carcinoma. J. Clin. Oncol. 23, 2903–2910.

- Chan, M., Hui, A., Yip, S., Ng, C., Lo, K., Tong, J., Chan, A., Cheung, H., et al., 2009. Progressive increase of genetic alteration in urinary bladder cancer by combined allelotyping analysis and comparative genomic hybridization. Int. J. Oncol. 34, 963–970
- Chapman, E., Harriden, P., Chambers, P., Johnston, C., Knowles, M., 2005. Comprehensive analysis of CDKN2A status in microdissected urothelial cell carcinoma reveals potential haploinsufficiency, a high frequency of homozygous codeletion and associations with clinical phenotype. Clin. Cancer Res. 11, 5740–5747.
- Chiou, H.Y., Chiou, S.T., Hsu, Y.H., Chou, Y.L., Tseng, C.H., Wei, M.L., Chen, C.J., 2001. Incidence of transitional cell carcinoma and arsenic in drinking water: a follow-up study of 8,102 residents in an arseniasis-endemic area in northeastern Taiwan. Am. J. Epidemiol. 153, 411–418.
- Chiu, H.F., Tsai, S.S., Yang, C.Y., 2007. Nitrate in drinking water and death from bladder cancer: an ecological case-control study in Taiwan. J. Toxicol. Environ. Health 70, 1000–1004.
- Christoph, F., Hinz, S., Kempkensteffen, C., Weikert, S., Krause, H., Schostak, M., Schrader, M., Miller, K., 2007. A gene expression profile of tumor suppressor genes commonly methylated in bladder cancer. J. Cancer Res. Clin. Oncol. 133, 343–340
- Christoph, F., Weikert, S., Kempkensteffen, C., Krause, H., Schostak, M., Miller, K., Schrader, M., 2006. Regularly methylated novel pro-apoptotic genes associated with recurrence in transitional cell carcinoma of the bladder. Int. J. Cancer 119, 1396–1402.
- Dyrskjøt, L., Ostenfeld, M., Bramsen, J., Silahtaroglu, A., Lamy, P., Ramanathan, R., et al., 2009. Genomic profiling of microRNAs in bladder cancer: miR-129 is associated with poor outcome and promotes cell death in vitro. Cancer Res. 69, 4851–4860.
- Dyrskot, L., Zieger, K., Real, F., Malats, N., Carrato, A., Hurst, C., Kotwal, S., Knowles, M., et al., 2007. Gene expression signatures predict outcome in non-muscle invasive bladder carcinoma: a multicenter validation study. Clin. Cancer Res. 13, 3545–3551
- Dyrskot, L., Zieger, K., Kruhoffer, M., Thykjaer, T., Jensen, J., Primdahl, H., Aiz, N., et al., 2005. A molecular signature in superficial bladder carcinoma predicts clinical outcome. Clin. Cancer Res. 11, 4029–4036.
- Ecke, T., Sachs, M., Lenk, S., Loening, S., Schlechte, H., 2008. TP53 gene mutations as an independent marker for urinary bladder cancer progression. Int. J. Mol. Med. 21, 655–661.
- Esteller, M., 2005. DNA methylation and cancer therapy: new developments and expectations. Curr. Opin. Oncol. 17, 55–60.
- Figueroa, J.D., Malats, N., Real, F.X., Silverman, D., Kogevinas, M., Chanock, S., et al., 2007. Genetic variation in the base excision repair pathway and bladder cancer risk. Hum. Genet. 121, 233–242.
- Fedewa, S.A., Soliman, A.S., Ismail, K., Hablas, A., Seifeldin, I.A., Ramadan, M., et al., 2009. Incidence of bladder cancer in the Nile delta region of Egypt. Cancer Epidemiol. 33, 176–181.
- Fontana, L., Fiori, M., Albini, S., Cifaldi, L., Giovinazzi, S., Forloni, M., Boldrini, R., Donfrancesco, A., et al., 2008. Antagomir-17-5p abolishes the growth of therapyresistant neuroblastoma through p21 and BIM. PLoS One 3, e2236.
- Franco, R., Schoneveld, O., Georgakilas, A., Panayiotidis, M., 2008. Oxidative stress, DNA methylation and carcinogenesis. Cancer Lett. 266, 6–11.
- Gallucci, M., Guadagni, F., Marzano, R., Leonardo, C., Merola, R., Sentinelli, S., Ruggeri, E., et al., 2005. Status of the p53, p16, RB1 and HER-2 genes and chromosomes 3, 7, 9 and 17 in advanced bladder cancer: correlation with adjacent mucosa and pathological parameters. J. Clin. Pathol. 58, 367–371.
- pathological parameters. J. Clin. Pathol. 58, 367–371.
  García-Closas, M., Malats, N., Real, F.X., Yeager, M., Welch, R., Silverman, D., et al., 2007. Large-scale evaluation of candidate genes identifies associations between VEGF polymorphisms and bladder cancer risk. PLoS Genet. 3, e29.
- Giannakakis, A., Coukos, G., Hatzigeorgiou, A., Sandaltzopoulos, R., Zhang, L., 2007. miRNA genetic alterations in human cancers. Expert Opin. Biol. Ther. 7, 1375–1386.
- Golka, K., Wiese, A., Assennato, G., Bolt, H.M., 2004. Occupational exposure and urological cancer. World J. Urol. 21, 382–391.
- Gottardo, F., Liu, C., Ferracin, M., Calin, G., Fassan, M., Bassi, P., Sevignani, C., et al., 2007. Micro-RNA profiling in kidney and bladder cancers. Urol. Oncol. 25, 387–392.
- Gouda, I., Mokhtar, N., Bilal, D., El-Bolkainy, T., El-Bolkainy, N.M., 2007. Bilharziasis and bladder cancer: a time trend analysis of 9843 patients. J. Egypt Natl. Canc. Inst. 19, 158–162.
- Hanke, M., Hoefig, K., Merz, H., Feller, A., Kausch, I., Jocham, D., Warnecke, J., Sczakiel, G., in press. A robust methodology to study urine microRNA as tumor marker: microRNA-126 and microRNA-182 are related to urinary bladder cancer. Urol. Oncol.
- Hirao, S., Hirao, T., Marsit, C., Hirao, Y., Schned, A., Devi-Ashok, T., Nelson, H., et al., 2005. Loss of heterozygosity on chromosome 9q and p53 alterations in human bladder cancer. Cancer 104, 1918–1923.
- Huang, C., Ke, Q., Costa, M., Shi, X., 2004. Molecular mechanisms of arsenic carcinogenesis. Mol. Cell. Biochem. 255, 57–66.
- Hurst, C., Tomlinson, D., Williams, S., Platt, F., Knowles, M., 2008. Inactivation of the Rb pathway and overexpression of both isoforms of E2F3 are obligate events in bladder tumours with 6p22 amplification. Oncogene 27, 2716–2727.
- Ichimi, T., Enokida, H., Okuno, Y., Kunimoto, R., Chiyomaru, T., Kawamoto, K., et al., 2009. Identification of novel microRNA targets based on microRNA signatures in bladder cancer. Int. J. Cancer 125, 345–352.
- Jemal, A., Siegel, R., Ward, E., Hao, Y., Xu, J., Thun, M., 2009. Cancer statistics. CA Cancer J. Clin. 59, 225–249.

- Kaldor, J.M., Day, N.E., Kittelmann, B., Pettersson, F., Langmark, F., Pedersen, D., 1995. Bladder tumours following chemotherapy and radiotherapy for ovarian cancer: a case-control study. Int. J. Cancer 27, 1-6.
- Koed, K., Wiuf, C., Christensen, L., Wikman, F., Zieger, K., Møller, K., von der Maase, H., Orntoft, T., 2005. High-density single nucleotide polymorphism array defines novel stage and location-dependent allelic imbalances in human bladder tumors. Cancer Res. 65, 34–45.
- Kim, E.J., Jeong, P., Quan, C., Kim, J., Bae, S.C., Yoon, S.J., Kang, J.W., et al., 2005. Genotypes of TNF-alpha, VEGF, hOGG1, GSTM1, and GSTT1: useful determinants for clinical outcome of bladder cancer. Urology 65, 70–75.
- Knight, A., Askling, J., Granath, F., Sparen, P., Ekbom, A., 2004. Urinary bladder cancer in Wagener's granulomatosis: risks and relation to cyclophosphamide. Ann. Rheum. Dis. 63, 1307–1311.
- Lin, T., Dong, W., Huang, J., Pan, Q., Fan, X., Zhang, C., Huang, L., 2009. MicroRNA-143 as a tumor suppressor for bladder cancer. J. Urol. 181, 1372-1380.
- Lopez-Abente, G., Aragones, N., Ramis, R., Hernandez-Barrera, V., Perez-Gomez, B., Escolar-Pujolar, A., 2006. Municipal distribution of bladder cancer mortality in Spain: possible role of mining and industry. BMC Public Health 6, 17.
- Lopez-Beltran, A., Alvarez-Kindelan, J., Luque, R., Blanca, A., Quintero, A., et al., 2008. Loss of heterozygosity at 9q32-33 (DBC1 locus) in primary non-invasive papillary urothelial neoplasm of low malignant potential and low-grade urothelial carcinoma of the bladder and their associated normal urothelium. J. Pathol. 215, 263-272.
- Maruyama, R., Toyooka, S., Toyooka, K., Harada, K., Virmani, A., Zochbauer-Muller, S., et al., 2001. Aberrant promoter methylation profile of bladder cancer and its relationship to clinicopathological features. Cancer Res. 61, 8659–8663.
- Mercatelli, N., Coppola, V., Bonci, D., Miele, F., Costantini, A., Guadagnoli, M., Bonanno, E., et al., 2008. The inhibition of the highly expressed miR-221 and miR-222 impairs the growth of prostate carcinoma xenografts in mice. PLoS One 3, e4029.
- Mundt, K.A., Birk, T., Burch, M.T., 2003. Critical review of the epidemiological literature on occupational exposure of perchloroethylene and cancer. Int. Arch. Occup. Environ. Health 76, 473–491.
- Olsson, A., Feber, A., Edwards, S., Te Poele, R., Giddings, I., Merson, S., Cooper, C., 2006. Role of E2F3 expression in modulating cellular proliferation rate in human bladder and prostate cancer cells. Oncogene 26, 1028–1037.
- Qin, S., Chen, X., Xu, X., Shou, J., Bi, X., Ji, L., Han, Y., Cai, Y., et al., 2006. Detection of chromosomal alterations in bladder transitional cell carcinomas from Northern China by comparative genomic hybridization. Cancer Lett. 238, 230–239.
- Romanenko, A., Kakehashi, A., Morimura, K., Wanibuchi, H., Wei, M., Vozianov, A., Fukushima, S., 2009. Urinary bladder carcinogenesis induced by chronic exposure to persistent low-dose ionizing radiation after Chernobyl accident. Carcinogenesis 30, 1821–1831.
- Romanenko, A., Morimura, K., Wanibuchi, H., Wei, M., Zaparin, W., Vinnichenko, W., Kinoshita, A., Vozianov, A., Fukushima, S., 2003. Urinary bladder lesions induced by persistent chronic low-dose ionizing radiation. Cancer Sci. 94, 328–333.
- Romanenko, A., Morimura, K., Wei, M., Zaparin, W., Vozianov, A., Fukushima, S., 2002. DNA damage repair in bladder urothelium after the Chernobyl accident in Ukraine. J. Urol. 168, 973–977.
- Romanenko, A., Morimura, K., Wanibuchi, H., Salim, E.I., Kinoshita, A., Kaneko, M., Vozianov, A., Fukushima, S., 2000. Increased oxidative stress with gene alteration in urinary bladder urothelium after the Chernobyl accident. Int. J. Cancer 866, 790–798.
- Rouissi, K., Ouerhani, S., Marrakchi, R., Ben Slama, M.R., Sfaxi, M., Ayed, M., Chebil, M., El Gaaied, A.B., 2009. Combined effect of smoking and inherited polymorphisms in arylamine N-acetyltransferase 2, glutathione S-transferases M1 and T1 on bladder cancer in a Tunisian population. Cancer Genet. Cytogenet. 190, 101–107.

- Saito, Y., Friedman, J., Chihara, Y., Egger, G., Chuang, J., Liang, G., 2009. Epigenetic therapy upregulates the tumor suppressor microRNA-126 and its host gene EGFL7 in human cancer cells. Biochem. Biophys. Res. Commun. 379, 726–731.
- Sánchez-Carbayo, M., 2003. Use of high-throughput DNA microarrays to identify biomarkers for bladder cancer. Clin. Chem. 49, 23–31.
- Sánchez-Carbayo, M., Socci, N., Lozano, J., Li, W., Charytonowicz, E., Belbin, T., Prystowsky, M., et al., 2003. Gene discovery in bladder cancer progression using cDNA microarrays. Am. J. Pathol. 163, 505–516.
- Sathyanarayana, U., Maruyama, R., Padar, A., Suzuki, M., Grossman, H., Czerniak, B., Gazdar, A., 2004. Molecular detection of nonivasive and invasive bladder tumor tissues and exfoliated cells by aberrant promoter methylation of laminin-5 encoding genes. Cancer Res. 64, 1425–1430.
- Schabath, M.B., Spitz, M.R., Grossman, H.B., Zhang, K., Dinney, C.P., Zheng, P.J., Wu, X.J., 2003. Genetic instability in bladder cancer assessed by the comet assay. J. Natl. Cancer Inst. 95, 540–547.
- Shariat, S., Kokunaga, H., Zhou, J., Kim, J., Ayala, G., Benedict, W., Lerner, S., 2004. p53, p21 pRB and p16 expression predict clinical outcome in cystectomy with bladder cancer. J. Clin. Oncol. 22, 1014–1024.
- Simon, R., Struckmann, K., Schraml, P., Wagner, U., Forster, T., Moch, H., Fijan, A., et al., 2002. Amplification pattern of 12q13-q15 genes (MDM2, CDK4, GLI) in urinary bladder cancer. Oncogene 21, 2476–2483.
- Song, D.K., Xing, D.L., Zhang, L.R., Li, Z.X., Liu, J., Qiao, B.P., 2009. Association of NAT2, GSTM1, GSTT1, CYP2A6, and CYP2A13 gene polymorphisms with susceptibility and clinicopathologic characteristics of bladder cancer in Central China. Cancer Detect. Prev. 32, 416–423.
- Stein, J., Lieskovsky, G., Cote, R., Groshen, S., Feng, A.C., Boyd, S., et al., 2001. Radical cystectomy in the treatment of invasive bladder cancer: long-term results in 1,054 patients. J. Clin. Oncol. 10, 666–675.
- Stewart, S., Cardinez, C., Richardson, L., Norman, L., Kaufmann, R., Pechacek, T., et al., 2008. Surveillance for cancers associated with tobacco use—United States, 1999–2004. MMWR Surveill. Summ. 57, 1–33.
- Tang, G., Tang, X., Mendu, V., Tang, X., Jia, X., Chen, Q., He, L., 2008. The art of microRNA: various strategies leading to gene silencing via an ancient pathway. Biochim. Biophys. Acta 1779, 655–662.
- Tanguay, C., Harvey, I., Houde, M., Srigley, R., Tetu, B., 2003. Leiomyosarcoma of the urinary bladder following cyclophosphamide therapy: report of two cases. Mod. Pathol. 16. 512–514.
- Veerla, S., Lindgren, D., Kvist, A., Frigyesi, A., Staaf, J., Persson, H., Liedberg, F., et al., 2009. MiRNA expression in urothelial carcinomas: important roles of miR-10a, miR-222, miR-125b, miR-7 and miR-452 for tumor stage and metastasis, and frequent homozygous losses of miR-31. Int. J. Cancer 124, 2236–2242.
- Villanueva, C., Fernandez, F., Malats, N., Grimalt, J., Kogevinas, M., 2003. Metaanalysis of studies on individual consumption of chlorinated drinking water and bladder cancer. J. Epidemiol. Commun. Health 57, 166–173.
- Wallerand, H., Bakkar, A.A., de Medina, S.G., Pairon, J.C., Yang, Y.C., Vordos, D., et al., 2005. Mutations in TP53, but not FGFR3, in urothelial cell carcinoma of the bladder are influenced by smoking: contribution of exogenous versus endogenous carcinogens. Carcinogenesis 26, 177–184.
- Wang, G., Zhang, H., He, H., Tong, W., Wang, B., Liao, G., Chen, Z., Du, C., in press. Up-regulation of microRNA in bladder tumor tissue is not common. Int. Urol. Nephrol.
- White-Gilbertson, S., Kasman, L., McKillop, J., Tirodkar, T., Lu, P., Voelkel-Johnson, C., 2009. Oxidative stress sentisizes bladder cancer cells to TRAIL mediated apoptosis by down-regulation anti-apoptotic proteins. J. Urol. 182, 1178–1185.
- Yuan, J.M., Chan, K.K., Coetzee, G.A., Castelao, J.E., Watson, M.A., Bell, D.A., Wang, R., Yu, M.C., 2008. Genetic determinants in the metabolism of bladder carcinogens in relation to risk of bladder cancer. Carcinogenesis 29, 1386–1393.
- Zeegers, M.P., Selen, R.F., Kleinjans, J.C., Goldbohm, R.A., van der Brandt, P.A., 2006. Nitrate intake does not influence bladder cancer risk: the Netherlands cohort study. Environ. Health Perspect. 114, 1527–1531.